

Available online at www.sciencedirect.com



RTICLE IN PRESS



Regulation of specific target genes and biological responses by estrogen receptor subtype agonists

Dale C Leitman¹, Sreenivasan Paruthiyil², Omar I Vivar¹, Elise F Saunier², Candice B Herber¹, Isaac Cohen², Mary Tagliaferri² and Terence P Speed^{3,4}

Estrogenic effects are mediated through two estrogen receptor (ER) subtypes, ER α and ER β . Estrogens are the most

commonly prescribed drugs to treat menopausal conditions, but by non-selectively triggering both ER α and ER β pathways in different tissues they can cause serious adverse effects. The different sizes of the binding pockets and sequences of their activation function domains indicate that ER α and ER β should have different specificities for ligands and biological responses that can be exploited for designing safer and more selective estrogens. ER α and ER β regulate different genes by binding to different regulatory elements and recruiting different transcription and chromatin remodeling factors that are expressed in a cell-specific manner. ER α -selective and ER β -selective agonists have been identified that demonstrate that the two ERs produce distinct biological effects. ER α and ER β agonists are a promising new approach for treating specific conditions associated with menopause.

Addresses

¹ Department of Nutritional Science and Toxicology, University of California, Berkeley, CA, USA

²Bionovo Inc., Emeryville, CA, USA

³ Department of Statistics, University of California, Berkeley, CA, USA ⁴ Division of Bioinformatics, The Walter and Eliza Hall Institute of Medical Research, Parkville, Victoria, Australia

Corresponding author: Leitman, Dale C (dale@leitmanlab.com)

Current Opinion in Pharmacology 2010, 10:1-8

This review comes from a themed issue on Endocrine and metabolic diseases Edited by Gary Firestone

1471-4892/\$ - see front matter Published by Elsevier Ltd.

DOI 10.1016/j.coph.2010.09.009

Introduction

Estrogens have important actions in non-reproductive tissues, including the brain, urogenital tract, and bone. Because of their actions in these tissues, estrogens have been used for over 50 years to prevent and treat a variety of conditions affecting postmenopausal women, including hot flashes, urogenital atrophy, and osteoporosis. Estrogens would be the clear drug of choice for treating menopausal symptoms if they did not cause some serious adverse effects. The most troublesome side-effect of estrogens is the increased risk of breast and endometrial cancer $[1^{\circ},2]$. Estrogens also increase blood clotting that can lead to venous thromboembolisms, and possibly strokes and heart disease, particularly in older women $[1^{\circ}]$.

Estrogens in hormone therapy (HT) were formulated long before there was a significant understanding of the mechanism of action of estrogens. The identification of ER α and ER β (Figure 1) and the crystal structures of their ligand binding domain (LBD), the discovery of a variety of coregulatory proteins involved in the genomic pathway and the demonstration of the nongenomic actions of estrogens [3,4] provide an extraordinary opportunity to design a new generation of estrogens that are safer and more selective. Estrogen receptor subtype agonists (ERSAs) [5,6°,7,8°,9,10°] have been identified (Figure 2) that might represent new classes of drugs to treat menopausal conditions. Here we will review ER α and ER β regulation of genes and the actions of several ERSAs and their potential clinical applications.

Differences in ER α and ER β are important for designing ERSAs

ERs are composed of three major modular domains; an A/ B domain, a DNA binding domain (DBD), and a LBD. Several features differ between ER α and ER β that might be important for designing ERSAs. First, the sizes of the ER α and ER β binding pocket for ligands are different, providing a structural basis for designing ligands that selectively bind to each ER. Second, the two activation function (AF-1 and AF-2) domains that are responsible for regulating gene transcription are located in the least homologous regions (Figure 1). The A/B domain containing AF-1 has only 17% homology, whereas the LBD that containing the AF-2 is 55% homologous. Differences in AF-1 and AF-2 could allow drugs to be designed that recruit different cofactors to ER α and ER β , thereby causing a different pattern of genes regulated.

ER α and ER β have distinct cellular actions, which provide a rationale for developing ERSAs. This has been demonstrated with microarrays that showed ER α and ER β regulate different genes [11,12,13°,14]. Only 40% of genes regulated by estradiol (E₂) in U2OS cells that express ER α were also regulated by ER β [12]. Furthermore,

www.sciencedirect.com

Current Opinion in Pharmacology 2010, 10:1-8

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/j.coph.2010.09.009

2 Endocrine and metabolic diseases

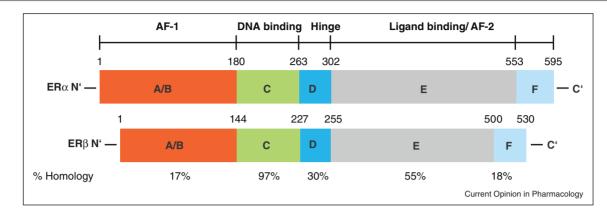


Figure 1

Comparison of the structures and homology between ER α and ER β . Human ER α contains 595 amino acids whereas ER β contains 530 amino acids. The DNA binding domains are nearly identical whereas the A/B domain and LBD, which contains AF-1 and AF-2, respectively, have the least homology.

ER α and ER β regulate different classes of genes suggesting that the two ERs have distinct physiological roles. Another feature that distinguishes ERB from ER α is that ER β regulates three classes of genes, whereas ER α regulates a single class of genes [15[•]]. U2OS cell lines stably transfected with a doxycycline-inducible ERa or ERB [15[•]] were used to measure the effects of unliganded ER in cells treated only with doxycycline or liganded-ER when cells were treated with both doxycycline and E₂. Unliganded ER α produced a small upregulation of only 1 gene and downregulation of 3 genes, whereas the addition of E_2 to doxycycline treated U2OS-ERa cells resulted in the activation of 518 genes and repression of 157 genes. These data indicate that ER α requires the ligand to regulate gene transcription in U2OS cells. By contrast, three classes of genes were regulated in U2OS-ERB cells. 453 genes were regulated by unliganded ERB (Class I genes). 258 genes were not regulated by unliganded ER β , but regulated by E_2 -bound ER β (Class II genes). 83 genes were regulated by unliganded ER β and potentiated by the addition of E₂ (Class III genes). The unliganded effect of ERB is mediated by AF-2, because it is lost when the ERB AF-2 is replaced by the ER α AF-2 [16]. These results demonstrate that intrinsic differences in AF-2 of ER α and ER β can lead to a different set of regulated genes regulated.

ER α and ER β regulate different genes by binding to distinct regulatory elements

A major question is how do ER α and ER β regulate different genes. The first step required for estrogens to regulate gene transcription involves the binding of ligand to the LBD. This causes a conformational change that allows the ligand-ER complex to bind to regulatory elements in target genes. ER α and ER β might regulate different genes by binding to different regulatory elements on target genes. To explore this possibility, ChIP-sequencing was performed in U2OS cells that express a stably transfected ER α or ER β to identify ER binding sites. 11,975 binding sites were found for ER β in response to E₂ [15°] and 15,947 binding sites for ER α (unpublished data). There was approximately a 30% overlap between ER α and ER β binding sites. Different ER α and ER β binding sites were also observed in MCF-7 cells [17°,18]. There were 4405 ER α and 1897 ER β binding sites, of which 1386 binding sites were common. These results demonstrate that many ER α and ER β binding sites are unique in U2OS and MCF-7 cells.

Tiling arrays [19,20[•],21] and ChIP-seq [15[•],22] studies demonstrated that many ER binding sites are more diverse and complex than the classical estrogen responsive element (ERE), requiring multiple different transcription factors for activity, such as AP1, FoxA1, and Sp1 [15,19,20,21–23]. The complexity is exemplified by the regulatory element in the NKG2E gene that requires a collaboration between c-jun, heat-shock factor 2, and CCAAT/enhancer-binding protein beta and a unique variant ERE for full activation by E₂ [24]. In MCF-7 cells T-cell factor and p53 motifs were present only in ERα binding sites [17[•]], whereas forkhead transcription factors and Sp1 sites were enriched in ERa and ERB sites, respectively [18]. These observations suggest that transcription factor binding elements are a major determinant of whether ER α or ER β will bind to a particular gene.

www.sciencedirect.com

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/j.coph.2010.09.009

Regulation of specific target genes and biological responses by estrogen receptor subtype agonists Leitman et al. 3

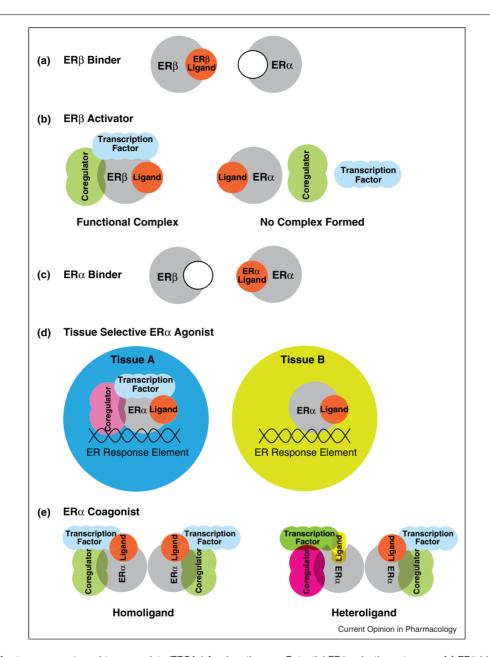


Figure 2

Potential classes of estrogen receptor subtype agonists (ERSAs) for drug therapy. Potential ER β -selective estrogens. (a) ER β binders (ERB-041) are estrogens that are selective because they bind to ER β with a much higher affinity than ER α . (b) ER β activators (MF101, liquiritigenin) bind to ER α and ER β with a similar affinity, and form a functional complex when bound to ER β (left panel), but not ER α (right panel). An ER β binder/activator (DPN) selectively binds to (a) and activates ER β (b). Potential ER α -selective agonists. (c) ER α binders (PPT) bind to ER α with a much higher affinity than ER β . (d) Tissue selective ER α agonists (Radix Glycyrrhiza and Radix Pueraria) form a functional transcription complex at response elements with ER α in some tissues (left panel), but not in other tissues (right panel). (e) A ligand such as E₂ binds to both ER α subunits that leads to the recruitment of coregulators and transcription factors (left panel). In the presence of an ER α coagonist (chalcone) E₂ binds to one subunit and the coagonist binds to the other subunit (right panel). The heteroliganded ER α could create a different conformation than the homoliganded ER α that leads to the recruitment of different coregulators and/or transcription factors.

$\text{ER}\alpha$ and $\text{ER}\beta$ regulate different genes by recruiting distinct coregulators and chromatin remodeling factors

Once the ER complex attaches to a regulatory element it functions as a docking site for the recruitment of

coregulatory proteins, and transcription and chromatin remodeling factors to form a large protein complex that regulates transcription [25,26]. Even if ER α and ER β bind to the same site they could regulate different genes because differences in their conformation might lead to

www.sciencedirect.com

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/j.coph.2010.09.009

the recruitment of different coregulatory proteins at the same genes. For example, liquiritigenin (LIQ) caused the recruitment of the coactivator, NCOA2 to the *CECR6*, *NKG2E*, and *NKD* genes in U2OS-ER β cells, but not in U2OS-ER α cells [9]. Furthermore, GIOT-4 has been identified as an ER β specific coactivator [27], whereas a member of the SWI/SNF chromatin remodeling complex, BAF57 selectively regulates ER α -mediated transcription [28].

Identification of three classes of $\text{ER}\beta$ -selective agonists

Multiple ER_β-selective agonists have been synthesized [10[•]]. 2,3-bis(4-hydroxyphenyl)-propionitrile (DPN) has 70-fold higher relative binding affinity and 170-fold higher relative potency in transfection assays with ERB compared to $ER\alpha$ [7]. Wyeth synthesized a number of ERβ-selective compounds [29]. ERB-041 has been the most studied. It has over a 200-fold greater selectivity for binding to ER β compared to ER α [6[•]]. In addition to synthetic compounds, a plant extract, MF101 contains ER β -selective agonists [8[•]], several of which have been identified, including liquiritigenin and nyasol [9,13[•]]. Based on binding and functional studies, we proposed that these compounds can be grouped into three classes [13[•]] (Figure 2). One class is represented by ERB-041 that is selective because it binds to $ER\beta$ at a much higher affinity than ER α (Figure 2a). We termed it an ER β binder. MF101, LIQ, and nysasol bind to both ERα and ER β similarly, but they only activate ER β [13[•]]. When these compounds bind to ER α they produce an inactive conformation that prevents ERa from forming a functional complex and recruiting coactivators [8,9] (Figure 2b). These are termed ERB activators. DPN is selective because it not only binds $ER\beta$ with higher affinity, but also more potently activates ERB than ER α . We termed it an ER β binder/activator. While most genes regulated by DPN, ERB-041, MF101, LIQ, and nyasol are the same, these three classes of ER β agonists regulate some different genes [13[•]]. Importantly, many genes regulated by these ERB agonists in U2OS-ERB cells are distinct from those regulated by E_2 . This observation is consistent with the finding that ERB binding sites are different when it is bound to ERB-041 compared to E_2 in MCF-7 cells [17[•]]. From these results, it can be expected that different classes of ERB agonists will produce different biological and clinical effects from one another and non-selective estrogens used in HT.

ER β -selective agonists for hot flashes

Estrogens are the most effective treatment for hot flashes. However, it is unclear if this effect is mediated through ER α , ER β , or both ERs. This has been difficult to address experimentally because of inadequate animal models to test drugs on spontaneous hot flashes. Most studies used rat models that measure tail skin temperature as a surrogate marker for hot flashes. In a morphineaddicted rat model two ERB-041 analogs were ineffective [30], whereas DPN was effective in another rat model [31]. A Phase II clinical trial with 217 postmenopausal women having moderate to severe hot flashes was conducted with the ER β -selective plant extract, MF101. After 12 weeks, there was a statistically significant median 11.9% reduction in hot flashes and a 67% reduction in night sweats in women treated with MF101 compared to those treated with placebo [32]. Taken together, these results suggest that ER β agonists might have beneficial effects on hot flash prevention.

$\text{ER}\beta$ -selective agonists for breast cancer prevention

Multiple studies showed that ERa mediates the proliferative effects of estrogens in breast cells. Anti-proliferative effects of ERB have been demonstrated in breast cancer cells [33,34]. In MCF-7 breast cancer cells, ERβ causes a G2 cell cycle arrest [34] by inhibiting the activity of cyclin dependent kinase 1 (CDK1) that is essential for cells to progress from G2 phase to mitosis. The major activator of CDK1 is cyclin B1. ERB inhibits the transcription of the cyclin B1 gene that leads to a reduction in cyclin B1 protein levels (submitted). CDK1 is inhibited by the tumor suppressor proteins, GADD45A and BTG2. $ER\beta$ binds to the promoter of these genes leading to increased transcription (submitted). Ultimately, the reduction in cyclin B1 and increased production of GADD45A and BTG2 leads to the inactivation of CDK1 and a G2 cell cycle arrest.

ERB-041 did not produce proliferative effects in the rat mammary gland [6[•]]. MF101 did not stimulate growth promoting genes, such as c-myc and cyclin D1 in MCF-7 cells [8[•]]. Furthermore, MF101 or LIQ did not increase MCF-7 cell tumor formation in mouse xenograft models $[8^{\circ},9]$. These results demonstrate that ER β agonists do not promote proliferation of normal mouse mammary epithelial and human breast cancer cells. ERB inhibits ERa-mediated activation of reporter genes in transfection assays [35], suggesting that one mechanism whereby $ER\beta$ exerts an anti-proliferative action is by interfering with the action of ER α . This was examined in MCF-7 cells that express ERa, ERB, or both ERs [17[•]]. These studies showed that ER α and ER β competed for the same genomic binding sites and that the presence of both ERs produced new binding sites for ER α and ER β homodimers, which probably leads to a different gene expression profile that is observed when the two ERs are coexpressed in cells [36]. These findings suggest that ER β agonists might be useful for preventing breast cancer by antagonizing the proliferative action of ER α .

$ER\beta$ -selective agonists for inflammatory diseases

One important action of estrogens that is relatively unappreciated is their anti-inflammatory effect. A number of

Current Opinion in Pharmacology 2010, 10:1–8

www.sciencedirect.com

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/ j.coph.2010.09.009

diseases during menopause have an inflammatory component to their pathogenesis. These conditions include osteoporosis, cardiovascular disease, Alzheimer's disease, obesity, and atrophic vaginitis. Estrogens in HT are very effective at preventing osteoporosis and atrophic vaginitis, but controversy exists regarding their effects on cardiovascular disease, obesity, and Alzheimer's disease. The anti-inflammatory action of ERB-041 has been examined in multiple inflammatory rodent models, including endometriosis, rheumatoid arthritis, inflammatory bowel disease, and sepsis [6,37,38]. These studies demonstrated that ERB-041 was very potent at blocking inflammation in these models and suggested that ERBselective agonists might be important drugs to treat a variety of disorders associated with inflammation. MF101 and synthetic ERB agonists, including ERB-041 are potent repressors of pro-inflammatory genes [8,39], indicating that estrogens can produce anti-inflammatory actions through ERB. The effects of ERB on inflammatory conditions associated with menopause, such as osteoporosis, obesity, cardiovascular disease, and atrophic vaginitis are unclear. ERB-041 did not prevent ovariectomy-induced bone loss or weight gain in rats [6[•]], suggesting that $ER\alpha$ mediates these effects. DPN decreases the size of infarcts in mouse hearts subjected to ischemia and reperfusion similar to E_2 [40]. This cardioprotective effect of DPN was abolished in ERB knockout mice [40]. These findings indicate that ERB agonists might be useful for preventing cardiovascular disease. Another possible clinical indication for $ER\beta$ agonists, where an anti-inflammatory effect could be therapeutic is atrophic vaginitis. Our pre-clinical studies with mice indicate that ER β agonists may play a role in the treatment of postmenopausal vaginal atrophy and dryness.

$\text{ER}\alpha$ is important for preventing osteoporosis, weight gain and insulin resistance

ER α is essential for preventing osteoporosis because a rare genetic mutation that inactivates ER α leads to severe osteoporosis in humans [41]. The observation that PPT, but not ERB-041 prevents bone loss in rats after ovariectomy provides additional evidence that ER α mediates the beneficial effects of estrogens in bone [6°,42]. ER α also probably mediates the beneficial effects of estrogens in adipose tissue and on insulin resistance, because ERKO mice have increased weight gain, greater adipose tissue, insulin resistance, and impaired glucose tolerance [43]. PPT prevents weight gain in rats and exerts antidiabetic effects by improving insulin sensitivity and glucose intolerance [44].

ER α -selective agonists

The major concern for developing ER α agonists is that they will cause cell proliferation and increase the risk of cancer. In fact, PPT stimulates the proliferation of HC11 mouse mammary epithelial cells [45] and increases uterine weight in rats [42]. These findings indicate that ER α selective binders (Figure 2c), like PPT might not be useful drugs for hormone therapy. Another strategy would be to design tissue selective ER α agonists that activate ER α in some tissues, such as the bone and adipose tissue, but not in the mammary gland and uterus (Figure 2d). An alternative strategy is to combine estrogens with other compounds that block the proliferative effects of estrogens in the mammary gland and uterus (Figure 2e). Progestins are effective at blocking the proliferative effects of estrogens in the uterus, but unfortunately they exacerbate the proliferative effects in the mammary gland.

Tissue selective $ER\alpha$ agonists prevent weight gain without promoting cell proliferation

It is well established that estrogens exert tissue-specific effects, but the mechanism is unclear. Tiling arrays identified 1090 ERa binding sites on chromosomes 1 and 6 in MCF-7 cells whereas 1137 ERa binding sites were found in U2OS cells [46[•]]. Only 172 ERa binding sites were common to both cell types. The cell-specific recruitment of ER α is mediated by the binding of the pioneer factor, FoxA1 that recognizes monomethylated and dimethylated histone H3. Once FoxA1 recognizes these methylated histones near an ER binding site it interacts with ER to open up chromatin structure and facilitate the recruitment of transcription factors leading to increased transcription [46[•]]. Because FoxA1 is expressed in MCF-7 cells, but not U2OS cells, the genes regulated by ER α are different [46[•]]. These findings suggest that it might be possible to design tissue selective ER α modulators that mimic the agonist activity of E₂ in some tissues, but not in other tissues.

We identified two plant extracts (PEs), Radix Glycyrrhiza and Radix Pueraria that behave as tissue selective ERa agonists (Figure 2d). These PEs activate ERa in transfection assays using an ERE upstream of the luciferase reporter and bind to purified $ER\alpha$ (In preparation). To test the effects of the PEs on weight loss, ovariectomized mice were fed a high fat diet (HFD). After the mice gained weight, they were treated orally for 6 weeks with the PEs separately while being maintained on the HFD. The vehicle treated control mice continued to gain weight, whereas the E₂-treated mice, which served as positive controls, lost 20.5% of their weight. The body weight and abdominal fat of both PE treated mice was significantly reduced to levels similar to mice treated with E₂. By contrast to E2 no significant proliferative effects were found in the mammary gland and uterus. While further characterization and studies are needed with the PEs these studies suggest that it might be possible to develop tissue selective ER α agonists that retain the beneficial effects mediated by ER α without promoting breast and endometrial cancer.

www.sciencedirect.com

Current Opinion in Pharmacology 2010, 10:1-8

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/ j.coph.2010.09.009

$\text{ER}\alpha$ coagonists change the gene expression profile and proliferative response of E_2

Another potential way to make estrogens safer for drug therapy is to add a second drug to alter the biological properties of estrogens after they interact with ER α . We screened plant extracts and found that a chalcone derivative dramatically changed the gene expression profile by E_2 in U2OS cells expressing ER α . We termed the chalcone an ER α coagonist (Figure 2e), because it was inactive by itself, but it caused E_2 to regulate genes that it did not activate in its absence and it potentiates the regulation of E_2 on some genes (In preparation). The coagonist blocked E₂-mediated proliferation of MCF-7 cells, suggesting that the coagonist changes the proliferative response of E₂ by causing ER α to regulate a different set of genes. While the mechanism of the coagonist is unclear, our studies suggest the possibility that it binds to $ER\alpha$ as heteroligand with one subunit binding to E_2 and the other subunit binding to the chalcone (Figure 2e, right panel). The combination of two different ligands bound to ER α simultaneously probably produces a different conformation than when $ER\alpha$ is bound to only E_2 (Figure 2e, left panel) or the chalcone. While the effects of the coagonist on E₂-mediated bone loss, weight loss, and mammary gland and endometrial cell proliferation in animals need to be investigated, it may be possible that coagonist compounds can alter the clinical responses to estrogens and make them safer.

Concluding remarks

Gene expression data, tiling arrays, and ChIP-seq data show that ER α and ER β regulate different genes by binding to distinct regulatory elements and interacting with different coactivators and transcription factors. Animal studies demonstrated that ERa-selective and ERβ-selective agonists produce different biological effects. Three classes of ERβ-selective agonists have been identified; ERB binder, ERB activator, and ERB binder/activator. ERβ-selective agonists might be clinically useful for preventing breast cancer and treating hot flashes and inflammatory conditions associated with menopause. Because the proliferative effects of estrogens are mediated through $ER\alpha$, the impetus to design ER α -selective agonists for clinical use has not been strong as for ER β -selective agonists. However, ER α is clearly important for preventing osteoporosis, weight gain, and insulin resistance. Tissue selective ERa agonists or ER α coagonists may provide a safer approach if proven to activate ER α in tissues that are beneficial, such as the bone and adipose tissue, but not the mammary gland and uterus. While many additional studies are needed to evaluate the safety and efficacy of ERa-selective and ER β -selective agonists they offer a new therapeutic approach for preventing and treating specific menopausal conditions.

Conflict of interest

O.I.V. and C.B.H. have nothing to declare. S.P., E.F.S., I.C., and M.T., are employees of Bionovo, Inc. D.C.L. and T.P.S. are on the Scientific Advisory Board of Bionovo, Inc. D.C.L. has received financial support for research from Bionovo, Inc.

Acknowledgement

We thank the National Center for Complementary and Alternative Medicine for funding our research.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- · of special interest
- 1. Writing Group for the Women's Health Initiative: Risks
- and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. *JAMA* 2002, 288:321–333.

This seminal study was the first randomized, placebo controlled study to evaluate the effects of hormone therapy on postmenopausal women. The study found that the risks of hormone therapy exceed the benefits.

- 2. Shang Y: Molecular mechanisms of oestrogen and SERMs in endometrial carcinogenesis. *Nat Rev Cancer* 2006, 6:360-368.
- Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, Tujague M, Strom A, Treuter E, Warner M et al.: Estrogen receptors: how do they signal and what are their targets. *Physiol Rev* 2007, 87:905-931.
- 4. Levin ER: Plasma membrane estrogen receptors. *Trends* Endocrinol Metab 2009, **20**:477-482.
- Stauffer SR, Coletta CJ, Tedesco R, Nishiguchi G, Carlson K, Sun J, Katzenellenbogen BS, Katzenellenbogen JA: Pyrazole ligands: structure-affinity/activity relationships and estrogen receptor-alpha-selective agonists. J Med Chem 2000, 43:4934-4947.
- 6. Harris HA, Albert LM, Leathurby Y, Malamas MS, Mewshaw RE,
- Miller CP, Kharode YP, Marzolf J, Komm BS, Winneker RC et al.: Evaluation of an estrogen receptor-beta agonist in animal models of human disease. Endocrinology 2003, 144:4241-4249.

models of human disease. Endocrinology 2003, **144**:4241-4249. This important study showed that a highly selective ER β agonist, ERB-041 had potent anti-inflammatory effects in multiple inflammation models and suggested that ER β agonists could be useful for treating inflammatory diseases.

- Meyers MJ, Sun J, Carlson KE, Marriner GA, Katzenellenbogen BS, Katzenellenbogen JA: Estrogen receptorbeta potency-selective ligands: structure-activity relationship studies of diarylpropionitriles and their acetylene and polar analogues. J Med Chem 2001, 44:4230-4251.
- 8. Cvoro A, Paruthiyil S, Jones JO, Tzagarakis-Foster C, Clegg NJ,
- Tatomer D, Medina RT, Tagliaferri M, Schaufele F, Scanlan TS et al.: Selective activation of estrogen receptor-beta transcriptional pathways by an herbal extract. Endocrinology 2007, 148:538-547.

This study demonstrated that plants contain ER β agonists, which do not stimulate the proliferation of human breast cancer cells or stimulate growth of the uterus in mice.

- Mersereau JE, Levy N, Staub RE, Baggett S, Zogric T, Chow S, Ricke WA, Tagliaferri M, Cohen I, Bjeldanes LF *et al.*: Liquiritigenin is a plant-derived highly selective estrogen receptor beta agonist. *Mol Cell Endocrinol* 2008, 283:49-57.
- 10. Minutolo F, Macchia M, Katzenellenbogen BS,
- Katzenellenbogen JA: Estrogen receptor beta ligands: recent advances and biomedical applications. *Med Res Rev* 2009. doi:10.1002/med.20186.

This important review summarizes the synthesis and biological properties of the ER β -selective compounds that have been developed.

Current Opinion in Pharmacology 2010, 10:1-8

www.sciencedirect.com

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/ j.coph.2010.09.009

Regulation of specific target genes and biological responses by estrogen receptor subtype agonists Leitman et al. 7

- Monroe DG, Getz BJ, Johnsen SA, Riggs BL, Khosla S, Spelsberg TC: Estrogen receptor isoform-specific regulation of endogenous gene expression in human osteoblastic cell lines expressing either ERalpha or ERbeta. J Cell Biochem 2003, 90:315-326.
- Kian Tee M, Rogatsky I, Tzagarakis-Foster C, Cvoro A, An J, Christy RJ, Yamamoto KR, Leitman DC: Estradiol and selective estrogen receptor modulators differentially regulate target genes with estrogen receptors {alpha} and {beta}. Mol Biol Cell 2004, 15:1262-1272.
- Paruthiyil S, Cvoro A, Zhao X, Wu Z, Sui Y, Staub RE, Baggett S, Herber CB, Griffin C, Tagliaferri M et al.: Drug and cell type-specific regulation of genes with different classes of estrogen receptor beta-selective agonists. PLoS ONE 2009, 4:e6271.

This study demonstrates that different ER β agonists regulate different genes from each other and from estradiol, suggesting that they will have different clinical effects.

- Stossi F, Barnett DH, Frasor J, Komm B, Lyttle CR, Katzenellenbogen BS: Transcriptional profiling of estrogenregulated gene expression via estrogen receptor (ER) alpha or ERbeta in human osteosarcoma cells: distinct and common target genes for these receptors. *Endocrinology* 2004, 145:3473-3486.
- Vivar OI, Zhao X, Saunier EF, Griffin C, Mayba OS, Tagliaferri M,
 Cohen I, Speed TP, Leitman DC: Estrogen receptor [beta] binds to and regulates three distinct classes of target genes. *J Biol Chem* 2010, 285:22059-22066.

This study demonstrates that there is a single class of genes regulated by ER α that requires the ligand, whereas ER β regulates three classes of genes, including genes regulated by unliganded ER β , genes regulated only by liganded ER β and genes regulated by both unliganded and liganded ER β .

- Levy N, Paruthiyil S, Zhao X, Vivar OI, Saunier EF, Griffin C, Tagliaferri M, Cohen I, Speed TP, Leitman DC: Unliganded estrogen receptor-beta regulation of genes is inhibited by tamoxifen. *Mol Cell Endocrinol* 2010, 315:201–207.
- Charn TH, Liu ET, Chang EC, Lee YK, Katzenellenbogen JA,
 Katzenellenbogen BS: Genome-wide dynamics of chromatin binding of estrogen receptors alpha and beta: mutual restriction and competitive site selection. *Mol Endocrinol* 2010, 24:47–59.

This important study shows that there is mutual competition for binding sites when both ER α and ER β are present and that presence of both receptors creates novel binding sites for ER α and ER β homodimers.

- Liu Y, Gao H, Marstrand TT, Strom A, Valen E, Sandelin A, Gustafsson JA, Dahlman-Wright K: The genome landscape of ERalpha- and ERbeta-binding DNA regions. Proc Natl Acad Sci USA 2008, 105:2604-2609.
- Carroll JS, Liu XS, Brodsky AS, Li W, Meyer CA, Szary AJ, Eeckhoute J, Shao W, Hestermann EV, Geistlinger TR *et al.*: Chromosome-wide mapping of estrogen receptor binding reveals long-range regulation requiring the forkhead protein FoxA1. *Cell* 2005, 122:33-43.
- Carroll JS, Meyer CA, Song J, Li W, Geistlinger TR, Eeckhoute J,
 Brodsky AS, Keeton EK, Fertuck KC, Hall GF et al.: Genome-wide
- Brousky AS, Rector EK, Pertuck KC, Hall GF et al.: Genome-wide analysis of estrogen receptor binding sites. Nat Genet 2006, 38:1289-1297.

This was an important study that first identified genome-wide $\mathsf{ER}\alpha$ binding sites and showed that ER binding sites are diverse and complex, requiring the interaction of multiple different transcription factors.

- Lupien M, Eeckhoute J, Meyer CA, Wang Q, Zhang Y, Li W, Carroll JS, Liu XS, Brown M: FoxA1 translates epigenetic signatures into enhancer-driven lineage-specific transcription. *Cell* 2008, 132:958-970.
- Welboren WJ, van Driel MA, Janssen-Megens EM, van Heeringen SJ, Sweep FC, Span PN, Stunnenberg HG: ChIP-Seq of ERalpha and RNA polymerase II defines genes differentially responding to ligands. *EMBO J* 2009, 28:1418-1428.
- Lin CY, Vega VB, Thomsen JS, Zhang T, Kong SL, Xie M, Chiu KP, Lipovich L, Barnett DH, Stossi F *et al.*: Whole-genome cartography of estrogen receptor alpha binding sites. *PLoS Genet* 2007, 3:e87.

- Levy N, Zhao X, Tang H, Jaffe RB, Speed TP, Leitman DC: Multiple transcription factor elements collaborate with estrogen receptor {alpha} to activate an inducible estrogen response element in the NKG2E gene. Endocrinology 2007, 148:3449-3458.
- Metivier R, Penot G, Hubner MR, Reid G, Brand H, Kos M, Gannon F: Estrogen receptor-alpha directs ordered, cyclical, and combinatorial recruitment of cofactors on a natural target promoter. *Cell* 2003, 115:751-763.
- Stanisic V, Lonard DM, O'Malley BW: Modulation of steroid hormone receptor activity. Prog Brain Res 2010, 181:153–176.
- Kouzu-Fujita M, Mezaki Y, Sawatsubashi S, Matsumoto T, Yamaoka I, Yano T, Taketani Y, Kitagawa H, Kato S: Coactivation of estrogen receptor beta by gonadotropin-induced cofactor GIOT-4. Mol Cell Biol 2009, 29:83-92.
- Garcia-Pedrero JM, Kiskinis E, Parker MG, Belandia B: The SWI/ SNF chromatin remodeling subunit BAF57 is a critical regulator of estrogen receptor function in breast cancer cells. *J Biol Chem* 2006, 281:22656-22664.
- Malamas MS, Manas ES, McDevitt RE, Gunawan I, Xu ZB, Collini MD, Miller CP, Dinh T, Henderson RA, Keith JC Jr et al.: Design and synthesis of aryl diphenolic azoles as potent and selective estrogen receptor-beta ligands. J Med Chem 2004, 47:5021-5040.
- Manas ES, Unwalla RJ, Xu ZB, Malamas MS, Miller CP, Harris HA, Hsiao C, Akopian T, Hum WT, Malakian K et al.: Structure-based design of estrogen receptor-beta selective ligands. J Am Chem Soc 2004, 126:15106-15119.
- Bowe J, Li XF, Kinsey-Jones J, Heyerick A, Brain S, Milligan S, O'Byrne K: The hop phytoestrogen, 8-prenylnaringenin, reverses the ovariectomy-induced rise in skin temperature in an animal model of menopausal hot flushes. *J Endocrinol* 2006, 191:399-405.
- Grady D, Sawaya GF, Johnson KC, Koltun W, Hess R, Vittinghoff E, Kristof M, Tagliaferri M, Cohen I, Ensrud KE: MF101, a selective estrogen receptor beta modulator for the treatment of menopausal hot flushes: a phase II clinical trial. *Menopause* 2009, 16:458-465.
- Strom A, Hartman J, Foster JS, Kietz S, Wimalasena J, Gustafsson JA: Estrogen receptor beta inhibits 17betaestradiol-stimulated proliferation of the breast cancer cell line T47D. Proc Natl Acad Sci USA 2004, 101:1566-1571.
- Paruthiyil S, Parmar H, Kerekatte V, Cunha GR, Firestone GL, Leitman DC: Estrogen receptor beta inhibits human breast cancer cell proliferation and tumor formation by causing a G2 cell cycle arrest. *Cancer Res* 2004, 64:423-428.
- 35. Hall JM, McDonnell DP: The estrogen receptor beta-isoform (ERbeta) of the human estrogen receptor modulates ERalpha transcriptional activity and is a key regulator of the cellular response to estrogens and antiestrogens. *Endocrinology* 1999, 140:5566-5578.
- Monroe DG, Secreto FJ, Subramaniam M, Getz BJ, Khosla S, Spelsberg TC: Estrogen receptor alpha and beta heterodimers exert unique effects on estrogen- and tamoxifen-dependent gene expression in human U2OS osteosarcoma cells. *Mol Endocrinol* 2005, 19:1555-1568.
- Cristofaro PA, Opal SM, Palardy JE, Parejo NA, Jhung J, Keith JC Jr, Harris HA: WAY-202196, a selective estrogen receptor-beta agonist, protects against death in experimental septic shock. *Crit Care Med* 2006, 34:2188-2193.
- Harris HA, Bruner-Tran KL, Zhang X, Osteen KG, Richard Lyttle C: A selective estrogen receptor-{beta} agonist causes lesion regression in an experimentally induced model of endometriosis. *Hum Reprod* 2005, 20:936-941.
- Cvoro A, Tatomer D, Tee MK, Zogovic T, Harris HA, Leitman DC: Selective estrogen receptor-agonists repress transcription of proinflammatory genes. J Immunol 2008, 180:630-636.
- 40. Lin J, Steenbergen C, Murphy E, Sun J: Estrogen receptor-beta activation results in S-nitrosylation of proteins involved in cardioprotection. *Circulation* 2009, **120**:245-254.

www.sciencedirect.com

Current Opinion in Pharmacology 2010, 10:1-8

Please cite this article in press as: Leitman DC, et al. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists, Curr Opin Pharmacol (2010), doi:10.1016/j.coph.2010.09.009

COPHAR-861; NO. OF PAGES 8

8 Endocrine and metabolic diseases

- 41. Grumbach MM, Auchus RJ: Estrogen: consequences and implications of human mutations in synthesis and action. *J Clin Endocrinol Metab* 1999, **84**:4677-4694.
- 42. Harris HA, Katzenellenbogen JA, Katzenellenbogen BS: Characterization of the biological roles of the estrogen receptors, ERalpha and ERbeta, in estrogen target tissues in vivo through the use of an ERalpha-selective ligand. *Endocrinology* 2002, **143**:4172-4177.
- Heine PA, Taylor JA, Iwamoto GA, Lubahn DB, Cooke PS: Increased adipose tissue in male and female estrogen receptor-alpha knockout mice. *Proc Natl Acad Sci USA* 2000, 97:12729-12734.
- 44. Lundholm L, Bryzgalova G, Gao H, Portwood N, Falt S, Berndt KD, Dicker A, Galuska D, Zierath JR, Gustafsson JA *et al*.:

The estrogen receptor {alpha}-selective agonist propyl pyrazole triol improves glucose tolerance in ob/ob mice; potential molecular mechanisms. *J Endocrinol* 2008, 199:275-286.

- Helguero LA, Faulds MH, Gustafsson JA, Haldosen LA: Estrogen receptors alfa (ERalpha) and beta (ERbeta) differentially regulate proliferation and apoptosis of the normal murine mammary epithelial cell line HC11. Oncogene 2005, 24:6605-6616.
- 46. Krum SA, Miranda-Carboni GA, Lupien M, Eeckhoute J, Carroll JS,
 Brown M: Unique ERalpha cistromes control cell type-specific and regulation Mol Endocrinol 2008 22:393-2406

gene regulation. *Mol Endocrinol* 2008, **22**:2393-2406. This important study demonstrated that the pioneer factor FoxA1 is important for cell-specific regulation of genes by estrogens.